# Caterpillars and moths

# Part I. Dermatologic manifestations of encounters with Lepidoptera

Eric W. Hossler, MD Danville, Pennsylvania

Caterpillars are the larval forms of moths and butterflies and belong to the order Lepidoptera. Caterpillars, and occasionally moths, have evolved defense mechanisms, including irritating hairs, spines, venoms, and toxins that may cause human disease. The pathologic mechanisms underlying reactions to Lepidoptera are poorly understood. Lepidoptera are uncommonly recognized causes of localized stings, eczematous or papular dermatitis, and urticaria. Part I of this two-part series on caterpillars and moths reviews Lepidopteran life cycles, terminology, and the epidemiology of caterpillar and moth envenomation. It also reviews the known pathomechanisms of disease caused by Lepidopteran exposures and how they relate to diagnosis and management. Part II discusses the specific clinical patterns caused by Lepidopteran exposures, with particular emphasis on groups of caterpillars and moths that cause a similar pattern of disease. It also discusses current therapeutic options regarding each pattern of disease. (J Am Acad Dermatol 2010;62:1-10.)

*Learning objectives:* After completing this learning activity, participants should be able to appropriately use current terminology of adverse reactions to caterpillars and moths, understand the epidemiology of these reactions, and use our current understanding of the pathologic mechanisms of these reaction patterns to guide treatment.

Key words: dermatitis; insect bites and stings; Lepidoptera; moths; urticaria.

aterpillars are the worm-like larval forms of Lepidoptera, the order of insects consisting of moths and butterflies. With an estimated 125,000 to 150,000 described species,<sup>1</sup> the Lepidoptera are one of the most prolific and diverse insect groups. In the United States alone there are approximately 13,000 known species, with 5000 found east of the Mississippi River.<sup>2</sup> Caterpillars and sometimes moths are an uncommon cause of disease in humans. Because these insects fall victim to many larger predators, they have developed irritating hairs, sharp spines, and various toxins to aid in their defense. Secondarily, these defenses occasionally can have impact on unwary humans. Myriad adverse reactions may result; these reactions have been

0190-9622/\$36.00

historically lumped into broad categories that have little clinical value and often are used interchangeably. Although the pathogenesis of adverse reactions to Lepidoptera is poorly understood, a number of different mechanisms appear to be at work. Understanding these pathways may help guide appropriate therapy.

# LIFE CYCLE AND TERMINOLOGY Key points

- Lepidoptera undergo four life stages: egg, caterpillar, pupa or chrysalis, and adult
- There has been confusion and overlap as to the terminology applied to adverse reactions to Lepidoptera
- Classifying reactions to Lepidoptera based on symptoms may be more useful than applying terms such as erucism and lepidopterism

All Lepidoptera are holometabolous (ie, there are four distinct life stages). Moths and butterflies represent the reproductive phase of Lepidoptera. After mating, they lay eggs from which caterpillars hatch. Caterpillars are the growing phase of Lepidoptera and feed primarily on plants. As

From the Department of Dermatology, Geisinger Medical Center, Danville.

Funding sources: None.

Conflicts of interest: The authors, editors, and peer reviewers have no relevant financial relationships.

Reprint requests: Eric W. Hossler, MD, Department of Dermatology, Geisinger Medical Center, 115 Woodbine Ln, Danville, PA 17822. E-mail: ewhossler@geisinger.edu.

<sup>© 2009</sup> by the American Academy of Dermatology, Inc. doi:10.1016/j.jaad.2009.08.060

caterpillars grow, they periodically outgrow their skin, which must be molted. The time between molts is called an instar. At the end of the growing phase, caterpillars enter a dormant phase called a pupa or chrysalis. Inside the pupa or chrysalis, the moth or butterfly develops and eventually hatches, completing the life cycle.<sup>1,2</sup>

**CAPSULE SUMMARY** 

caterpillars.

The order Lepidoptera comprises moths,

butterflies, and their worm-like larvae,

Caterpillars, and less commonly moths,

cause localized stings, eczematous

membrane irritation, or, rarely, life-

The pathophysiologic mechanisms of

but may involve irritant reactions,

hypersensitivity reactions, and toxic

these reactions are poorly understood

threatening hemorrhage.

envenomation.

may bear irritating hairs or spines that

eruptions, urticarial reactions, mucous

There is significant confusion as to the terminology referring to adverse events from contact with Lepidoptera. Erucism has been defined as either any reaction from caterpillars<sup>3,4</sup> or any reaction limited to the skin from caterpillars or moths.<sup>5</sup> Lepidopterism can mean any reaction to caterpillars or moths,<sup>4,6,7</sup> refer only to reactions from contact with scales or hairs from adult moths or butterflies,<sup>5</sup> or refer only to cases with systemic signs and symptoms, with or without cutaneous findings, resulting from contact with any lepidopteran

source.<sup>8,9</sup> The word "erucism" derives from the Latin *eruca*, meaning caterpillar,<sup>10</sup> while "lepidopterism" stems from the Greek words lepis, meaning scale or flake, and *pteron*, meaning wing.<sup>11</sup> Strictly speaking, erucism should refer to any reaction from caterpillars and lepidopterism to reactions from moths or butterflies. Because reactions to both larval and adult Lepidoptera can cause a variety of either cutaneous and/or systemic symptoms, classifying reactions into erucism or lepidopterism is only of academic interest. I find it easier to avoid use of either term, and instead classify reactions based on predominant symptoms (eg, urticaria induced by contact with processionary caterpillars [genus Thaumetopoea]). In this manner, the clinician can continue to use familiar descriptive terms, such as urticarial or eczematous, and this approach will help both with diagnosis and treatment.

# EPIDEMIOLOGY

## Key points

- Documentation of caterpillar and moth reactions is scarce in the medical literature, and likely underrepresents the true number of cases
- Epidemics of erucism and lepidopterism are facilitated by natural abundance,

introduction of species into unnatural habitats, and, in some species, wind dispersion of larvae or setae

- Artificial illumination combined with natural abundance may congregate offensive species of moths, such as *Hylesia* and *Euproctis*
- A large proportion of caterpillar reactions

## are reported in children

Despite the diversity and nearly worldwide distribution of Lepidoptera, there are few species with clear documentation of adverse reactions in humans. Reasons for this are manifold and may include general medical disinterest, predominantly mild and self-limited reactions from Lepidoptera, difficulty in the accurate identification of offending species, and the occurrence of cases in tropical areas where medical reporting is either more difficult or less

prioritized.<sup>12</sup> The number of adverse lepidopteran exposures is difficult to quantify. No studies have been performed in a controlled setting. However, of 94,552 bites and stings annually reported by phone to poison control centers in the United States, only 2094 (2.2%) were reportedly related to caterpillar exposure.<sup>13</sup> Considering that most reactions are mild and self-limited, these numbers likely underestimate the true number of adverse reactions.

Human-Lepidoptera interactions are infrequent. Therefore, adverse events in humans typically occur in only one or a few exposed individuals. However, several species of Lepidoptera are prone to seasonal abundance, leading to increased exposure frequency and "epidemics" of cutaneous or systemic symptoms. This is well known in the northeast United States, where gypsy moth caterpillars (Lymantria dispar) defoliate large tracts of forest each summer. Epidemics of dermatitis from this caterpillar have been reported in Massachusetts, Connecticut, and Pennsylvania.<sup>14-16</sup> Outbreaks of the Douglas-fir tussock moth caterpillar (Orgvia pseudotsugata) in the Pacific Northwest have caused several epidemics of papular urticaria amongst loggers,<sup>17,18</sup> and more recently in a Boy Scout camp in New Mexico.<sup>19</sup> In Venezuela, *Hylesia* moths may be so prevalent that schools and shops are closed early, and farmers and fisherman are unable to work

because of a fear of rash and incapacitating itch.<sup>20</sup> The following species are also known to periodically outbreak: puss caterpillars (*Megalopyge opercularis*),<sup>21-23</sup> buck moth caterpillars (*Hemileuca maia*),<sup>24</sup> range caterpillars (*H oliviae*),<sup>25</sup> several *Euproctis* species,<sup>26-29</sup> and several species of processionary caterpillars (*Thaumetopoea*).<sup>30-32</sup> Temporary reduction in natural parasites was blamed for the abundance of *Hylesia alinda* moths, which caused an epidemic of dermatitis in Cozumel, Mexico, in 1989.<sup>33</sup> Introduced species, such as the gypsy moth, gum leaf skeletonizer moth (*Uraba lugens*), and stinging nettle moth (*Darna pallivita*) may benefit from a lack of natural predators, facilitating overabundance.

An additional problem is that some offending species have the ability to disseminate themselves widely. Newly hatched gypsy moth caterpillars, which may be more allergenic than mature larvae, are capable of wind dispersal by means of a silken thread, a behavior called "ballooning."<sup>15,34,35</sup> First instar larvae of the closely related Douglas-fir tussock moth are also capable of airborne dissemination in this manner.<sup>17,36</sup>

Caterpillar setae of some species are easily detached from larvae and can be widely dispersed by winds, causing dermatitis or ophthalmia nodosa.<sup>37</sup> This phenomenon has been documented with the oak processionary caterpillar (Tprocessionea), 30,32,38 pine processionary caterpillars (T pityocampa),<sup>39</sup> mistletoe browntail moth (Euproctis edwardsi),<sup>40</sup> E flava,28 and Hylesia.41 Garments hung on clotheslines may collect airborne setae and cause dermatitis when the clothes are worn.<sup>28,42</sup> Outbreaks may be massive: larval abundance, dry weather, and strong winds contributed to a 1972 outbreak in Shanghai, China, in which an estimated 500,000 cases of dermatitis were caused by airborne setae from the caterpillar of the Asian mulberry tussock moth (E flava).<sup>28</sup>

Although caterpillars cause the vast majority of adverse events from lepidopteran exposures, adult moths may also cause adverse reactions. When attracted to artificial lighting, the irritating setae from these moths may cause irritant or allergic reactions. The most well recognized moth reaction is Caripito itch, which is caused by setae from female moths of the genus Hylesia. Dinehart et al<sup>41</sup> reported that 34 of 35 crewman on an oil tanker were continuously affected with a pruritic eruption stemming from the setae of dead moths and moth parts aboard their ship that had been docked overnight in Caripito, Venezuela, 3 weeks earlier. Similarly, 54 of 55 crewman aboard an oil tanker docked at the port of Caripito were affected in the report by Zaias et al.<sup>43</sup> Another offensive moth

species is *Euproctis bipunctapex*. In 1990, 141 inhabitants of a public housing estate in Singapore suffered from a papular urticarial eruption caused by these moths; they had been attracted to the high-rise's lighting.<sup>44</sup>

Caterpillar and moth exposures are reported more frequently in children. Of the cases of caterpillar exposure reported to US Poison Centers, between 51.6% and 57% occurred in persons 18 years of age or younger.<sup>45,46</sup> Other series report that between 24% and 30% of exposures occur in children who are less than 6 years old.<sup>13,47</sup> In addition to these reports, a number of papers have independently reported a disproportionate number of children affected because of contact with the following species: the gum leaf skeletonizer (*Ulugens*),  $^{48}$  buck moth (H maia),<sup>24</sup> white-stemmed gum moth collesi),<sup>49</sup> (Chelepteryx Euproctis similis,<sup>50</sup> Lasiocampa quercus,<sup>50</sup> Hylesia metabus,<sup>51</sup> H alinda,<sup>33</sup> and Lonomia obliqua.<sup>52</sup> An outbreak of dermatitis and respiratory distress in a German kindergarten was blamed on an infestation of oak processionary caterpillars (T processionea) in nearby oak trees.<sup>30</sup> The reasons for this pediatric predominance are unclear, but may be related to frequent outdoor activity and increased curiosity. Derraik<sup>48</sup> postulated that the bright coloration of some caterpillars, such as the gum leaf skeletonizer (Ulugens), may be attractive to children, resulting in increased direct contact.<sup>48</sup> An alternative hypothesis is that caterpillar and moth exposures are not actually more common in children, but that a greater proportion of exposures are reported, perhaps due to parental concern.

### **PATHOGENESIS**

# **Key points**

- Some caterpillars bear setae and/or spines that may be directly irritating or possess venoms or toxins
- Although most moths are harmless, female *Hylesia* moths bear hollow spines
- Histamine has been extracted from several species and may play a role in human reactions
- Browntail moth (*Euproctis chrysorrhea*) caterpillar extracts have shown a wide variety of enzymatic properties
- Patch testing to caterpillar setae has shown an immediate hypersensitivity reaction, delayed-type hypersensitivity, or both
- Processionary caterpillars (genus *Thaumetopoea*) cause primarily a type I hypersensitivity reaction



**Fig 1.** Close view of setae from a gypsy moth caterpillar (*Lymantria dispar*).

- *Lonomia* caterpillars produce protein toxins that cause consumptive coagulopathy and fibrinolysis
- Ophthalmia nodosa may involve an immediate toxic response, followed by a foreign body granulomatous reaction
- Dendrolimiasis and pararamose both involve granulomatous inflammation involving joints

Almost all exposures to toxic Lepidoptera or their products are caused by either (1) direct contact with allergenic or irritating Lepidoptera parts, such as hairs or scales, or (2) stinging spines that may contain venoms. Most reactions are from accidental exposure; however, occupational exposure may occur in persons who rear or work with Lepidoptera,<sup>7,53</sup> and cases of dermatitis from commercial silkworm (*Bombyx mori*) cocoons and textiles made from silk have been reported. These occupational exposures have been documented as eczematous dermatitis<sup>54</sup> and contact urticaria.<sup>55</sup> The allergen in raw silk is unknown.<sup>56</sup> Even more uncommon are moths that intentionally bite humans and feed on blood or tears.<sup>3,57</sup>

#### Spines and setae

Caterpillars have developed an immense array of cutaneous appendages designed to repel would-be attackers or predators. One type of appendage is the hair-like seta, arising singly or in large bunches from



**Fig 2.** Close view of spines from the saddleback caterpillar (*Acharia stimulea*).

the integument of the caterpillar (Fig 1). Spines are more robust multicellular processes that are contiguous with the integument (Fig 2).<sup>1,3,58,59</sup> Spines are relatively fixed, and cause adverse reactions only when the insect comes in direct contact with human skin. In contrast, caterpillar setae may be detachable and easily rubbed off, even becoming airborne in some instances, and can be incorporated into the structure of the cocoon, ostensibly for pupal protection.<sup>3,59</sup> Female moths of the genus Hylesia transfer setae from their abdomen onto their egg mass, which may provide protection from ants or larger predators.<sup>60</sup> Contact with setae from Hylesia cocoons, egg masses, or other fomites may cause adverse reaction even without direct contact with the caterpillar. Setae from the oak processionary caterpillar (T processionea) are environmentally stable for at least 1 year, causing symptoms long after caterpillars would normally be found.<sup>32</sup>

Spines and setae come in myriad forms; Mullen<sup>3</sup> described and illustrated seven types of setae and four types of spines of medical importance. Both setae and spines may cause mechanical irritation or contain substances that possess histamine or other irritating substances, trigger the release of histamine, or have other enzymatic actions.<sup>3,61,62</sup> Some spines and setae do not cause mechanical irritation and symptoms appear attributable only to venoms or toxins. One author found that setae of various caterpillars (species not identified) had no direct irritating properties when inserted into the skin after they were rinsed with water or alcohols, arguing that only the chemicals carried by the setae are able to instigate adverse reactions.<sup>63</sup>

Although adult moths are usually harmless, moths of the genus *Hylesia* are well known for the pruritic rash that follows exposure. Female *Hylesia* moths bear hollow-tipped spines attached to gland-like cells on the abdomen (Fig 3). Moths of the African genus *Anaphe* bear similar spines<sup>64</sup> and have been reported to cause dermatitis similar to *Hylesia*.<sup>3,60,65</sup>



**Fig 3.** Spines from abdomen of female *Hylesia lineata*. **A**, Abdominal hairs (scales) from a female *H lineata*. A mix of these hairs forms the felt around the egg mass. **B**, Short urticating hairs (scales). Note barbs on lower ends. **C**, Modified felt-forming ends and barbs of long abdominal hairs (scales). (Reprinted with permission from Janzen DH. Natural history of *Hylesia lineata* in Santa Rosa National Park, Costa Rica. J Kansas Entomol Soc 1984;57:490-514.)

*Euproctis bipunctapex*<sup>44</sup> and *E flava*<sup>28,29,66</sup> moths may also cause dermatitis. Unlike *Hylesia*, *Euproctis* moths do not bear their own setae; instead, the caterpillar incorporates its setae into the cocoon when it pupates, and the moth picks up setae from the cocoon as it emerges.<sup>7,44,50</sup> Rarely, tibial spurs on larger moths can be strong enough to penetrate human skin and cause localized stings, dermatitis, or urticaria.<sup>64,67</sup> Finally, adult moths of the genus *Calyptra* have a stiff, barbed proboscis that can be used to penetrate intact mammalian skin in order to feed on blood.<sup>3,57,68</sup>

#### Chemical toxins and irritants

Considering the sheer number of species of Lepidoptera, it is not surprising that the chemical

makeup of toxins and irritants is so diverse. Histamine may play a role in the symptoms of many species, and it has been isolated in setae from caterpillars of the genus *Dirphia*,<sup>69</sup> setae of the gypsy moth (*L dispar*),<sup>70</sup> whole caterpillars of the browntail moth (Euproctis chrysorrhea) and the Japanese tea tussock moth (*E pseudoconspersa*),<sup>71-73</sup> and the venom of the gum leaf skeletonizer (Ulugens).<sup>59</sup> A histamine-like substance was found in the hairs of Spilosoma lutea.74 One group found histamine in hairs from Hylesia moths<sup>75</sup> and found that intradermal injections of Hylesia moth extract cause wheals that were abated by the administration of diphenhydramine but not indomethacin.<sup>75</sup> However, others have failed to demonstrate histamine in setae from at least one Hylesia species (H metabus).<sup>76,77</sup>

Other chemicals isolated from Lepidoptera include acetylcholine, found in caterpillar setae, moths, and eggs of the great tiger moth (*Arctia caja*).<sup>74,78</sup> Formic acid, typically found in ants, has been found in the secretions of several Notodontid caterpillars. One Notodontid species, the variable oakleaf caterpillar (*Lochmaeus manteo*), has caused skin blistering after brief contact.<sup>79</sup> In a rat model, extracts from *Dirphia* caterpillar setae were able to cause pleuritis that was reduced by pretreatment with dexamethasone, rofecoxib, pyrilamine, and sodium diclofenac. The authors concluded that extracts caused tissue damage by inducing histamine, proinflammatory products of cyclooxygenase, and nitric oxide.<sup>76</sup>

More complex enzymatic compounds may play a role in the dermatitis caused by several species. Proteins with trypsin-like activity and vasodegenerative and fibrinolytic effects have been identified in Hylesia moths.<sup>20,80</sup> Megalopyge urens venom has direct hemolytic and proteolytic activity but lacked both histamine and acetylcholine.<sup>81</sup> Euproctis caterpillars have been shown to harbor a host of enzymes that serve as potential irritants. Aqueous solutions made from larval hairs of browntail moth caterpillars (*E chrysorrhea*) have trypsin- and chymotrypsin-like properties and fibrinolytic, proteolytic, hemolytic, and anticomplement activity.<sup>61,72</sup> These extracts were able to consume complement, initiate histamine release, and generate plasmin from plasminogen.<sup>82</sup> Serine proteases, including kallikrein, have been found in the spicule venoms of the browntail moth and the closely related E subflava.<sup>83</sup> Phospholipase A and esterase are also present in both of these species and have been theorized to be responsible for the cutaneous reactions.<sup>72,82,84</sup> Numerous studies have shown the ability of browntail moth setal extracts to cause spherocytosis.<sup>82,85,86</sup> The relevance of this last finding is unknown.

Two species of Lonomia caterpillars, L obliqua and Lachelous, contain toxins that cause potentially fatal coagulation defects. Despite the clinical similarity of the hemorrhagic diathesis caused by envenomation, the toxin mechanisms for these two species appear to differ greatly. Caterpillars of L achelous contain several novel toxins ("Lonomins") that activate several hematologic pathways, including direct fibrinolysis, prothrombin activation, degradation of factor XIII, and factor Xa-like activity.52,87,88 Conversely, caterpillars of L obliqua contain two procoagulant toxins: Losac (Lonomia obliqua Stuartfactor activator; an activator of factor X) and Lopap (Lonomia obliqua prothrombin activator protease; an activator of prothrombin).<sup>52,89,90</sup> In contrast to the direct fibrinolysis caused by Lonomins from L achelous, envenomation by Lobliqua results in consumptive coagulopathy with secondary fibrinolysis.88,89 Lopap may be the major toxin; infusion of Lopap into mice causes a similar hemorrhagic diathesis, and an antivenin directed against Lopap is effective in reversing the coagulation defects.<sup>90</sup> Despite the differences in venoms and venom actions, both caterpillars cause a similar clinical picture of hemorrhagic diathesis, fibrinolysis, and "unclottable" blood, resulting in potentially fatal cutaneous, mucosal, visceral, and intracranial bleeding, with or without renal failure. Recently, caterpillar venom from Cerodirphia speciosa, a related species of the same subfamily (Hemileucinae), was found to have two proteins similar to Lonomia venom.91 Hemorrhage has not been reported after stings from this species.

#### Hypersensitivity reactions

In addition to mechanical irritation, venoms, and toxins, Lepidoptera are capable of causing hypersensitivity reactions in susceptible individuals. Some species cause immediate hypersensitivity reactions, others cause delayed-type hypersensitivity, and some appear capable of causing both. The following paragraphs summarize what is currently published in the literature. Understanding and prompt recognition of these reactions may help direct therapy.

There is evidence of a type 1 hypersensitivity to several species. Intradermal injections causing immediate wheal-and-flare reactions have been demonstrated with *Hylesia* moth and egg extracts and browntail moth (*E chrysorrhea*) and Japanese tea tussock moth (*E pseudoconspersa*) setal extracts. Preheating of the browntail moth extract reduced or eliminated these responses.<sup>86</sup> Scratch testing with gypsy moth caterpillars, cast caterpillar skins, and egg mass hairs caused wheal-and-flare reactions in 15 or 17 US Department of Agriculture Forest Service personnel who had been working with gypsy

moths.<sup>92</sup> Prick testing with Douglas-fir tussock moth (*Opseudotsugata*) larvae, cocoons, shed larval hairs, adults, and egg masses showed a wheal and flare reaction in all participants who had previously reported reactions to caterpillar exposure.<sup>93</sup>

Prick testing with extracts from processionary caterpillars (genus Thaumetopoea), which are known to cause urticarial reactions, cause significantly higher rates of positive testing in individuals with previous exposure to caterpillars when compared to those without contact.<sup>62,94-96</sup> In addition, immunoglobulin E (to hair extracts) has been found in the sera of forest workers exposed to pine processionary caterpillars (T pityocampa)<sup>39</sup> and in almost all patients who report urticaria, angioedema, or bronchial asthma after exposure to this caterpillar.<sup>96,97</sup> It appears that last-instar larvae of pine processionary caterpillars are the most allergenic and that allergenicity increases with each molt.<sup>98</sup> Finally, a protein named "Thaumetopoein" has been isolated from T pityocampa that acts directly on mast cells, causing immunoglobulin E-independent degranulation.99

Patch testing with caterpillar or moth setae has shown the presence of an immediate hypersensitivity, delayed-type hypersensitivity, or both. Hellier and Warin<sup>50</sup> patch tested the forearms of healthy volunteers with pieces of three species of caterpillars: Eriogaster lunestris, Spilosoma lubricipeda, and Euproctis similis. All those tested to Euproctis had marked pruritus and erythema within 48 hours, and six of 10 subjects tested with Eriogaster or Spilosoma reacted within 48 hours, the rest having a more mild response after the patches were removed at 48 hours.<sup>50</sup> Euproctis caterpillars may cause both immediate and delayed-type hypersensitivity reactions. Patch testing with setae from browntail moth caterpillars (E chrysorrhea) revealed erythema and edema within 5 hours, often progressing to vesiculation at 72 hours.<sup>86</sup> Patch testing with setal extracts from the Japanese tea tussock moth (E pseudoconspersa) showed both immediate and delayed-type reactions.<sup>73</sup> Patch testing with gypsy moth (*L dispar*) caterpillar hairs caused delayed papulovesicular reactions in eight of eight patients with a history of dermatitis caused by gypsy moths, while only one of 11 healthy controls reacted.<sup>100</sup> Patch tests using Douglas-fir tussock moth (O pseudotsugata) larvae, cocoons, shed larval hairs, adults, and egg masses showed primary irritant reaction with erythema occurring within 45 minutes of application and vesiculation at 24 hours.<sup>93</sup> Closed patch testing using female Hylesia moths or egg masses caused erythema beyond the patch test site within 15 minutes and vesiculation within hours.<sup>77</sup> Taken together, this

information lends credence to the theory that more than one mechanism of hypersensitivity may be at work, and that the cutaneous response may vary between individuals as well as species of Lepidoptera.

#### Other reactions

The mechanisms of ophthalmia nodosa are poorly understood. Both barbed and unbarbed hairs<sup>101</sup> cause immediate unilateral chemosis, which can progress to liquefactive necrosis and hypopyon acutely, and later can develop into a granulomatous reaction. Caterpillar hairs are frequently demonstrated within these granulomas,<sup>102-104</sup> suggesting a foreign body response. In many cases, however, the setae are never removed and the foreign hairs appear to be tolerated.<sup>102,103,105</sup>

Dendrolimiasis and pararamose (caused by exposure to Dendrolimus and Premolis semirufa caterpillars, respectively) are similar reactions that are both characterized by prominent arthritis in association with pruritic dermatitis. The mechanisms of these reactions are poorly understood, but both processes involve granuloma formation, often with bristle fragments embedded in periosteum, synovial membrane, or articular cartilage.<sup>106-108</sup> Dias and de Azevedo<sup>108</sup> found that setae from *P* semirufa were able to penetrate down to mouse perichondria, periosteum, tendon sheaths, and synovial bursae, causing granulomatous inflammation.<sup>108</sup> Huang<sup>109</sup> suggested that dendrolimiasis may be caused by allergic reaction, toxin envenomation, or secondary infection. Additional research is needed in this area.

#### REFERENCES

- 1. Gullan PJ, Cranston P. The insects: an outline of entomology, 3rd ed. London: Wiley-Blackwell; 2004.
- Wagner DL. Caterpillars of eastern North America: a guide to identification and natural history. Princeton, NJ: Princeton University Press; 2005.
- Mullen GR. Moths and butterflies (Lepidoptera). In: Mullen GR, Durden LA, editors. Medical and veterinary entomology. San Diego, CA: Academic Press; 2002. p. 364.
- Scoble MJ. The lepidoptera. New York: Oxford University Press; 1992.
- 5. Goddard J. Physician's guide to arthropods of medical importance. 2nd ed. Philadelphia: CRC Press; 1996.
- Elston DM. Bites and stings. In: Bolognia JL, Jorizzo JL, Rapini RP, Callen JP, Horn TD, Mancini AJ, editors. Dermatology, 2nd ed, vol. 1. New York: Elsevier; 2008. p. 1303.
- Wirtz RA. Allergic and toxic reactions to non-stinging arthropods. Ann Rev Entomol 1984;29:47-69.
- Diaz JH. The epidemiology, diagnosis, and management of caterpillar envenoming in the southern US. J La State Med Soc 2005;157:153-7.
- Diaz JH. The evolving global epidemiology, syndromic classification, management, and prevention of caterpillar envenoming. Am J Trop Med Hyg 2005;72:347-57.

- 10. Traupman JC. The Bantam new college Latin and English dictionary. 2nd ed. New York: Bantam Books; 1995.
- Crane GR. Perseus digital library 4.0. Available at: http://www.perseus.tufts.edu/hopper/. Accessed February 16, 2009.
- 12. Rosen T. Caterpillar dermatitis. Dermatol Clin 1990;8:245-52.
- Langley RL. Animal bites and stings reported by United States Poison Control Centers, 2001-2005. Wilderness Environ Med 2008;19:7-14.
- Tuthill RW, Canada AT, Wilcock K, Etkind PH, O'Dell TM, Shama SK. An epidemiologic study of gypsy moth rash. Am J Public Health 1984;74:799-803.
- 15. Anderson JF, Furniss WE. Epidemic of urticaria associated with first-instar larvae of the gypsy moth (Lepidoptera: Lymantriidae). J Med Entomol 1983;20:146-50.
- Centers for Disease Control and Prevention. Rash illness associated with gypsy moth caterpillars—Pennsylvania. MMWR Morb Mortal Wkly Rep 1982;31:169-70.
- Press E, Googins JA, Poareo H, Jones K. Health hazards to timber and forestry workers from the Douglas fir tussock moth. Arch Environ Health 1977;32:206-10.
- Hoover AH, Nelson E. Skin symptoms attributed to tussock moth infestation. Cutis 1974;13:597.
- 19. Redd JT, Voorhees RE, Torok TJ. Outbreak of lepidopterism at a Boy Scout camp. J Am Acad Dermatol 2007;56:952-5.
- Benaim-Pinto C, Pernia-Rosales B, Rojas-Peralta R. Dermatitis caused by moths of *Hylesia* genus (Lepidoptera, Saturniidae) in northeastern states of Venezuela: I. Bioecology of *Hylesia metabus* (Cramer). Clinical features of lepidopterism determined by this species. Am J Contact Derm 1991;2:213.
- McGovern JP, Barkin GD, McElhenney TR, Wende R. *Megalopyge opercularis*: observations of its life history, natural history of its sting in man, and report of an epidemic. JAMA 1961;175:1155-8.
- 22. Micks DW. Clinical effects of the sting of the "puss caterpillar" (*Megalopyge opercularis* S & A) on man. Tex Rep Biol Med 1952;10:399-405.
- 23. Bishopp FC. The puss caterpillar and the effects of its sting on man. United States Department of Agriculture Department Circulation 1923;288:1-4.
- 24. Walker RB, Thomas T, Cupit D, Giaquinto-Shreves J. An epidemic of caterpillar sting dermatitis in a rural West Virginia community. W V Med J 1993;89:58-60.
- Tuskes PM, Tuttle JP, Collins MM. The wild silk moths of North America: a natural history of the Saturniidae of the United States and Canada. Ithaca, NY: Cornell University Press; 1996.
- 26. Ferguson DC. Noctuoidea. Lymantriidae, vol. 22-2. London: E.W. Classey; 1978.
- 27. Blair CP. The browntail moth, its caterpillar and their rash. Clin Exp Dermatol 1979;4:215-22.
- De-Long S. Mulberry tussock moth dermatitis. A study of an epidemic of unknown origin. J Epidemiol Community Health 1981;35:1-4.
- 29. Tsutsumi C. A histological study of the development of the urticating spicules of the Far Eastern urticating moth, *Euproctis flava* Bremer (Lepidoptera: Lymantriidae). Jpn J Med Sci Biol 1958;11:443-53.
- Gottschling S, Meyer S, Dill-Mueller D, Wurm D, Gortner L. Outbreak report of airborne caterpillar dermatitis in a kindergarten. Dermatology 2007;215:5-9.
- Hesler LS, Logan TM, Benenson MW, Moser C. Acute dermatitis from oak processionary caterpillars in a U.S. military community in Germany. Mil Med 1999;164:767-70.
- 32. Maier H, Spiegel W, Kinaciyan T, Krehan H, Cabaj A, Schopf A, et al. The oak processionary caterpillar as the cause of an

epidemic airborne disease: survey and analysis. Br J Dermatol 2003;149:990-7.

- Centers for Disease Control and Prevention. Moth-associated dermatitis—Cozumel, Mexico. MMWR Morb Mortal Wkly Rep 1990;39:219-20.
- Liebhold S. Gypsy moth in North America. Available at: http://www.fs.fed.us/ne/morgantown/4557/gmoth/. Accessed April 11, 2008.
- 35. McManus ML. The role of behavior in the dispersal of newly hatched gypsy moth larvae. Available at: http://www.fs.fed. us/ne/newtown\_square/publications/research\_papers/pdfs/ scanned/OCR/ne\_rp267.pdf. Accessed March 14, 2008.
- Wickman BE, Manson RR, Trostle GC. Douglas-fir tussock moth. Available at: http://www.na.fs.fed.us/spfo/pubs/fidls/ tussock/fidl-tuss.htm. Accessed March 14, 2008.
- Bishop JW, Morton MR. Caterpillar-hair keratoconjunctivitis. Am J Ophthalmol 1967;64:778-9.
- Spiegel W, Maier H, Maier M. A non-infectious airborne disease. Lancet 2004;363:1438.
- Werno J, Lamy M, Vincendeau P. Caterpillar hairs as allergens. Lancet 1993;342:936-7.
- Balit CR, Ptolemy HC, Geary MJ, Russell RC, Isbister GK. Outbreak of caterpillar dermatitis caused by airborne hairs of the mistletoe browntail moth (*Euproctis edwardsi*). Med J Aust 2001;175:641-3.
- Dinehart SM, Archer ME, Wolf JE Jr, McGavran MH, Reitz C, Smith EB. Caripito itch: dermatitis from contact with Hylesia moths. J Am Acad Dermatol 1985;13:743-7.
- Dunlop K, Freeman S. Caterpillar dermatitis. Australas J Dermatol 1997;38:193-5.
- Zaias N, Ioannides G, Taplin D. Dermatitis from contact with moths (genus Hylesia). JAMA 1969;207:525-7.
- Ooi PL, Goh KT, Lee HS, Goh CL. Tussockosis: an outbreak of dermatitis caused by tussock moths in Singapore. Contact Derm 1991;24:197-200.
- Litovitz TL, Smilkstein M, Felberg L, Klein-Schwartz W, Berlin R, Morgan JL. 1996 annual report of the American Association of Poison Control Centers toxic exposure surveillance system. Am J Emerg Med 1997;15:447-500.
- 46. Bronstein AC, Spyker DA, Cantilena LR Jr, Green J, Rumack BH, Heard SE. 2006 annual report of the American Association of Poison Control Centers' National Poison Data System (NPDS). Clin Toxicol (Phila) 2007;45:815-917.
- 47. Everson GW, Chapin JB, Normann SA. Caterpillar envenomations: a prospective study of 112 cases. Vet Hum Toxicol 1990;32:114-9.
- Derraik JG. Three students exposed to Uraba lugens (gum leaf skeletoniser) caterpillars in a West Auckland school. N Z Med J 2007;120:U2656.
- 49. Balit CR, Geary MJ, Russell RC, Isbister GK. Clinical effects of exposure to the white-stemmed gum moth (*Chelepteryx collesi*). Emerg Med Australas 2004;16:74-81.
- 50. Hellier FF, Warin RP. Caterpillar dermatitis. Br Med J 1967;2: 346-8.
- Rodriguez-Morales AJ, Arria M, Rojas-Mirabal J, Borges E, Benitez JA, Herrera M, et al. Lepidopterism due to exposure to the moth *Hylesia metabus* in northeastern Venezuela. Am J Trop Med Hyg 2005;73:991-3.
- 52. Carrijo-Carvalho LC, Chudzinski-Tavassi AM. The venom of the *Lonomia* caterpillar: an overview. Toxicon 2007;49: 741-57.
- Wirtz RA. Occupational allergies to arthropods—documentation and prevention. Bull Entomol Soc Am 1980;26:356-60.
- Inoue A, Ishido I, Shoji A, Yamada H. Textile dermatitis from silk. Contact Dermatitis 1997;37:185.

- 55. Rudzki E. Contact urticaria from silk. Contact Dermatitis 1977; 3:52.
- Rietschell RL, Fowler JF, eds. Fisher's Contact Dermatitis, 5th Edition.; 2001
- Banziger H. Skin-piercing blood-sucking moths II: studies on a further 3 adult *Calyptra* [Calpe] sp. (Lepid., Noctuidae). Acta Trop 1979;36:23-37.
- Romoser WS. Introduction to arthropods: structure, function and development. In: Idridge BF, Edman JD, editors. Medical entomology: a textbook on public health and veterinary problems caused by arthropods. New York: Kluwer Academic Publishers; 2000. p. 13.
- Southcott R. Moths and butterflies. In: Covacevich J, Davie P, Pearn J, editors. Toxic plants and animals: a guide for Australia, 1st ed. Brisbane, Australia: Queensland Museum; 1987. p. 243.
- Rodriguez J, Hernandez JV, Fornes L, Lundberg U, Arocha-Pinango CL, Osborn F. External morphology of abdominal setae from male and female *Hylesia metabus* adults (Lepidoptera: Saturniidae) and their function. Fla Entomol 2004; 87:30-6.
- de Jong MC. Dermatitis caused by the larva of the brown-tail moth, *Euproctis chrysorrhoea* L. (Lymantriidae). Dermatologica 1977;154:310-1.
- 62. Vega JM, Moneo I, Armentia A, Fernández A, Vega J, De La Fuente R, et al. Allergy to the pine processionary caterpillar (*Thaumetopoea pityocampa*). Clin Exp Allergy 1999;29: 1418-23.
- Goldman L, Sawyer F, Levine A, Goldman J, Goldman S, Spinanger J. Investigative studies of skin irritations from caterpillars. J Invest Dermatol 1960;34:67-79.
- 64. Rothschild M, Reichstein T, von Euw J, Aplin R, Harman RR. Toxic lepidoptera. Toxicon 1970;8:293-9.
- 65. Bowles DE, Swaby JA. Field guide to venomous and medically important invertebrates affecting military operations: identification, biology, symptoms, treatment. Available at: http://www.afpmb.org/pubs/Field\_Guide/field\_guide.htm. Accessed October 6, 2008.
- Mills RG. Observations on a series of case of dermatitis caused by a Liparid moth, *Euproctis flava*. Chin Med J 1923; 37:351-71.
- 67. Rothschild M. Some observations on the garden tiger moth, Arctia caja L. Proc Royal Ent Soc Lond 1957;11:1.
- Zaspel VS, Kononenko VS, Goldstein PZ. Another blood feeder? Experimental feeding of a fruit-piercing moth species on human blood in the Primorye territory of far eastern Russia (Lepidoptera: Noctuidae: Calpinae). J Insect Behav 2007;20:437-51.
- Valle JR, Picarelli ZP, Prado JL. Histamine content and pharmacological properties of crude extracts from setae of urticating caterpillars. Arch Int Pharmacodyn Ther 1954;98:324-34.
- Shama SK, Etkind PH, Odell TM, Canada AT, Finn AM, Soter NA. Gypsy-moth-caterpillar dermatitis. N Engl J Med 1982; 306:1300-1.
- 71. Hall-Smith PJ, Graham P. Beware the furry caterpillar. Clin Exp Dermatol 1980;5:261-2.
- de Jong MC, Bleumink E. Investigative studies of the dermatitis caused by the larva of the brown-tail moth, *Euproctis chrysorrhoea* L. (Lepidoptera, Lymantriidae). III. Chemical analysis of skin reactive substances. Arch Dermatol Res 1977;259:247-62.
- 73. Natsuaki M. Immediate and delayed-type reactions in caterpillar dermatitis. J Dermatol 2002;29:471-6.

- Frazer JFD. The cause of urtication produced by larval hairs of *Arctia caja* (L.). Proc R Entomol Soc London 1965;40:96-100.
- Dinehart SM, Jorizzo JL, Soter NA, Noppakun N, Voss WR, Hokanson JA, et al. Evidence for histamine in the urticating hairs of Hylesia moths. J Invest Dermatol 1987;88:691-3.
- Lunardelli A, Leite CE, Pires MG, de Oliveira JR. Extract of the bristles of *Dirphia* sp. increases nitric oxide in a rat pleurisy model. Inflamm Res 2006;55:129-35.
- 77. Benaim-Pinto C, Pernia-Rosales B, Rojas-Peralta R. Dermatitis caused by moths of *Hylesia* genus (Lepidoptera, Saturniidae) in northeastern states of Venezuela: II. Biochemistry and immunoallergy of substances responsible for dermal lesions. Am J Contact Dermat 1992;3:5-15.
- Morley J, Schachter M. Acetylcholine in non-nervous tissues of some lepidoptera. J Physiol 1963;168:706-15.
- Kearby WH. Variable oakleaf caterpillar larvae secrete formic acid that causes skin lesions (Lepidoptera: Notodontidae). J Kansas Entomol Soc 1975;48:280-2.
- Lundberg U, Salazar V, Tovar M, Rodriguez J. Isolation and partial characterization of proteins with vasodegenerative and proinflammatory properties from the egg-nests of *Hylesia metabus* (Lepidoptera: Saturniidae). J Med Entomol 2007;44:440-9.
- Ardao MI, Sosa Perdomo C, Pellaton MG. Venom of the Megalopyge urens (Berg) caterpillar. Nature 1966;209:1139-40.
- de Jong MC, Bleumink E. Investigative studies of the dermatitis caused by the larva of the brown-tail moth, *Euproctis chrysorrhoea* L. (Lepidoptera, Lymantriidae). IV. Further characterization of skin reactive substances. Arch Dermatol Res 1977;259:263-81.
- Bleumink E, de Jong MC, Kawamoto F, Meyer GT, Kloosterhuis AJ, Slijper-Pal IJ. Protease activities in the spicule venom of *Euproctis* caterpillars. Toxicon 1982;20:607-13.
- de Jong MC, Kawamoto F, Bleumink E, Kloosterhuis AJ, Meijer GT. A comparative study of the spicule venom of *Euproctis* caterpillars. Toxicon 1982;20:477-85.
- Tyzzer EE. The pathology of the brown-tail moth dermatitis. J Med Res 1907;16:43-64.
- de Jong MC, Bleumink E, Nater JP. Investigative studies of the dermatitis caused by the larva of the brown-tail moth (*Euproctis chrysorrhoea* Linn.) I. Clinical and experimental findings. Arch Dermatol Res 1975;253:287-300.
- Chan K, Lee A, Onell R, Etches W, Nahirniak S, Bagshaw SM, et al. Caterpillar-induced bleeding syndrome in a returning traveller. CMAJ 2008;179:158-61.
- 88. Zannin M, Lourenco DM, Motta G, Dalla Costa LR, Grando M, Gamborgi GP, et al. Blood coagulation and fibrinolytic factors in 105 patients with hemorrhagic syndrome caused by accidental contact with *Lonomia obliqua* caterpillar in Santa Catarina, southern Brazil. Thromb Haemost 2003;89:355-64.
- Fritzen M, Flores MP, Reis CV, Chudzinski-Tavassi AM. A prothrombin activator (lopap) modulating inflammation, coagulation and cell survival mechanisms. Biochem Biophys Res Commun 2005;333:517-23.
- Chudzinki-Tavassi AM, Carrijo-Carvalho LC. Biochemical and biological properties of *Lonomia obliqua* bristle extract. J Venom Anim Toxins incl Trop Dis 2006;12:159-71.
- Shevchenko A, de Sousa MM, Waridel P, Bittencourt ST, de Sousa MV, Shevchenko A. Sequence similarity-based proteomics in insects: characterization of the larvae venom of the Brazilian moth *Cerodirphia speciosa*. J Proteome Res 2005;4: 862-9.

- 92. Etkind PH, Odell TM, Canada AT, Shama SK, Finn AM, Tuthill R. The gypsy moth caterpillar: a significant new occupational and public health problem. J Occup Med 1982;24:659-62.
- Perlman F, Press E, Googins JA, Malley A, Poarea H. Tussockosis: reactions to Douglas fir tussock moth. Ann Allergy 1976;36:302-7.
- 94. Vega J, Vega JM, Moneo I, Armentia A, Caballero ML, Miranda A. Occupational immunologic contact urticaria from pine processionary caterpillar (*Thaumetopoea pityocampa*): experience in 30 cases. Contact Dermatitis 2004;50:60-4.
- 95. Vega JM, Moneo I, Armentia A, Vega J, De la Fuente R, Fernandez A. Pine processionary caterpillar as a new cause of immunologic contact urticaria. Contact Dermatitis 2000;43: 129-32.
- Fuentes Aparicio V, Zapatero Remon L, Martinez Molero MI, Alonso Lebreros E, Beitia Mazuecos JM, Bartolome Zavala B. Allergy to pine processionary caterpillar (*Thaumetopoea pity-ocampa*) in children. Allergol Immunopathol 2006;34:59-63.
- Fuentes Aparicio V, de Barrio Fernández M, Rubio Sotés M, Rodríguez Paredes A, Martínez Molero MI, Zapatero Remón L, et al. Non-occupational allergy caused by the pine processionary caterpillar (*Thaumetopoea pityocampa*). Allergol Immunopathol 2004;32:69-75.
- Rebollo S, Moneo I, Vega JM, Herrera I, Caballero ML. Pine processionary caterpillar allergenicity increases during larval development. Int Arch Allergy Immunol 2002;128:310-4.
- Lamy M, Pastureaud MH, Novak F, Ducombs G, Vincendeau P, Maleville J, et al. Thaumetopoein: an urticating protein from

the hairs and integument of the pine processionary caterpillar (*Thaumetopoea pityocampa* Schiff., Lepidoptera, Thaumetopoeidae). Toxicon 1986;24:347-56.

- Beaucher WN, Farnham JE. Gypsy-moth-caterpillar dermatitis. N Engl J Med 1982;306:1301-2.
- Arora R, Gupta AK, Chaturvedi KU, Bhatnagar A. Ophthalmia nodosa: due to unbarbed hairs. J Pediatr Ophthalmol Strabismus 1994;31:104-6.
- 102. Corkey JA. Ophthalmia nodosa due to caterpillar hairs. Br J Ophthalmol 1955;39:301-6.
- Cadera W, Pachtman MA, Fountain JA, Ellis FD, Wilson FM 2nd. Ocular lesions caused by caterpillar hairs (ophthalmia nodosa). Can J Ophthalmol 1984;19:40-4.
- 104. Watson PG, Sevel D. Ophthalmia nodosa. Br J Ophthalmol 1966;50:209-17.
- 105. Horng CT, Chou PI, Liang JB. Caterpillar setae in the deep cornea and anterior chamber. Am J Ophthalmol 2000;129: 384-5.
- 106. Lawson JP, Liu YM. Pinemoth caterpillar disease. Skeletal Radiol 1986;15:422-7.
- 107. Costa RM, Atra E, Ferraz MB, da Silva NP, de Souza JM, Batista Júnior J, et al. "Pararamose": an occupational arthritis caused by lepidoptera (*Premolis semirufa*). An epidemiological study. Rev Paul Med 1993;111:462-5.
- 108. Dias LB, de Azevedo MC. Pararama, a disease caused by moth larvae: experimental findings. Pathol Bull 1973;7:9-14.
- 109. Huang DZ. Dendrolimiasis: an analysis of 58 cases. J Trop Med Hyg 1991;94:79-87.